



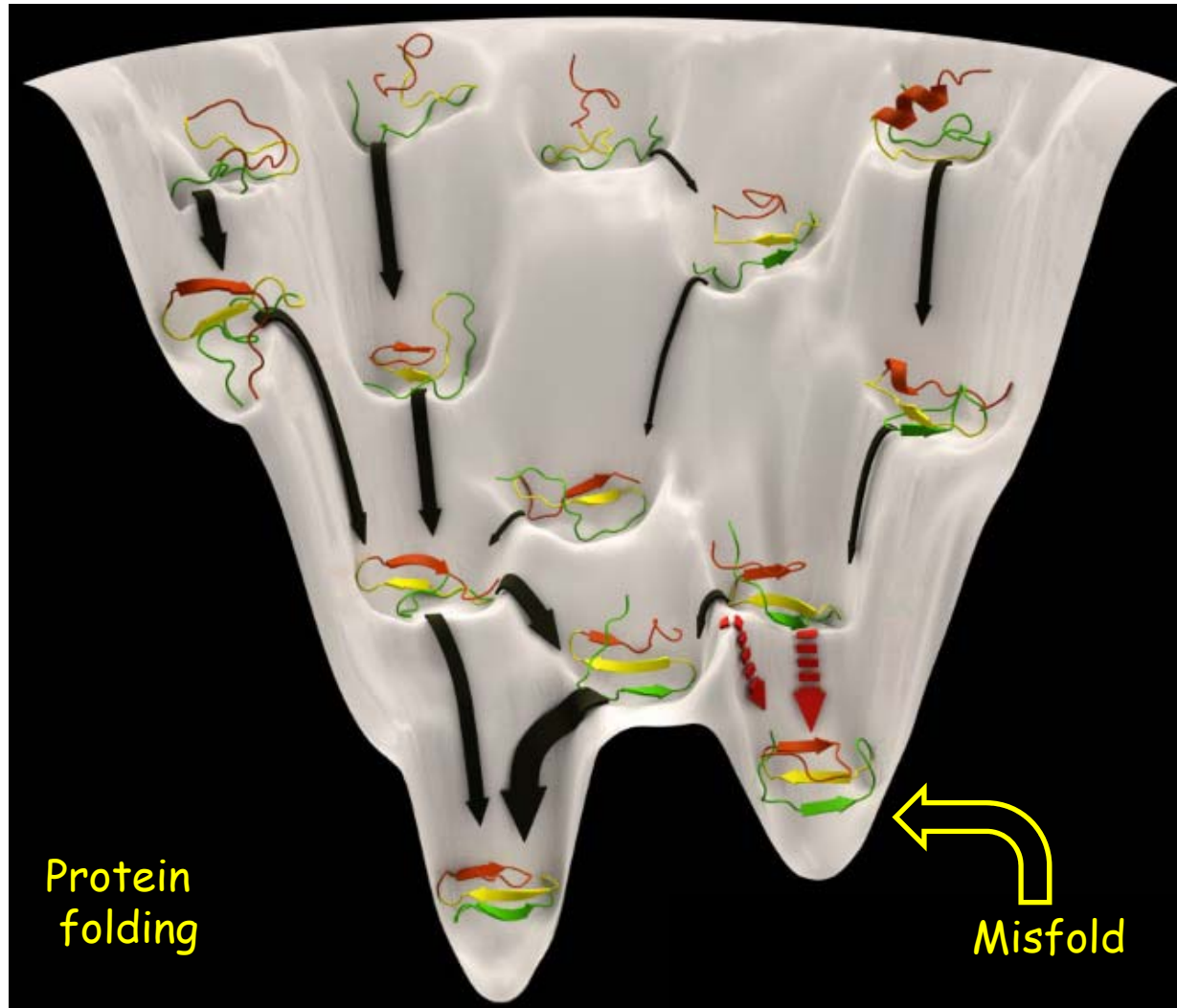
Ultrastable AFM:
Improved stability, precision,
& bandwidth for bioAFM

The image is a 3D schematic of an Atomic Force Microscope (AFM) probe. The probe is a long, thin, light blue structure with a sharp tip. It is shown in a perspective view, with its base on a dark blue surface. The probe is illuminated by several green and red laser beams. A red line graph is visible on the right side of the image, and a green line graph is visible on the left side. The probe is shown in two positions: one where it is just above the surface and another where it is in contact with the surface. The contact point is highlighted with a yellow glow. The surface is dark blue and has a textured appearance, suggesting a biological or material surface. The background is black.

Thomas Perkins, JILA

Protein folding: a 50-year old problem in biology

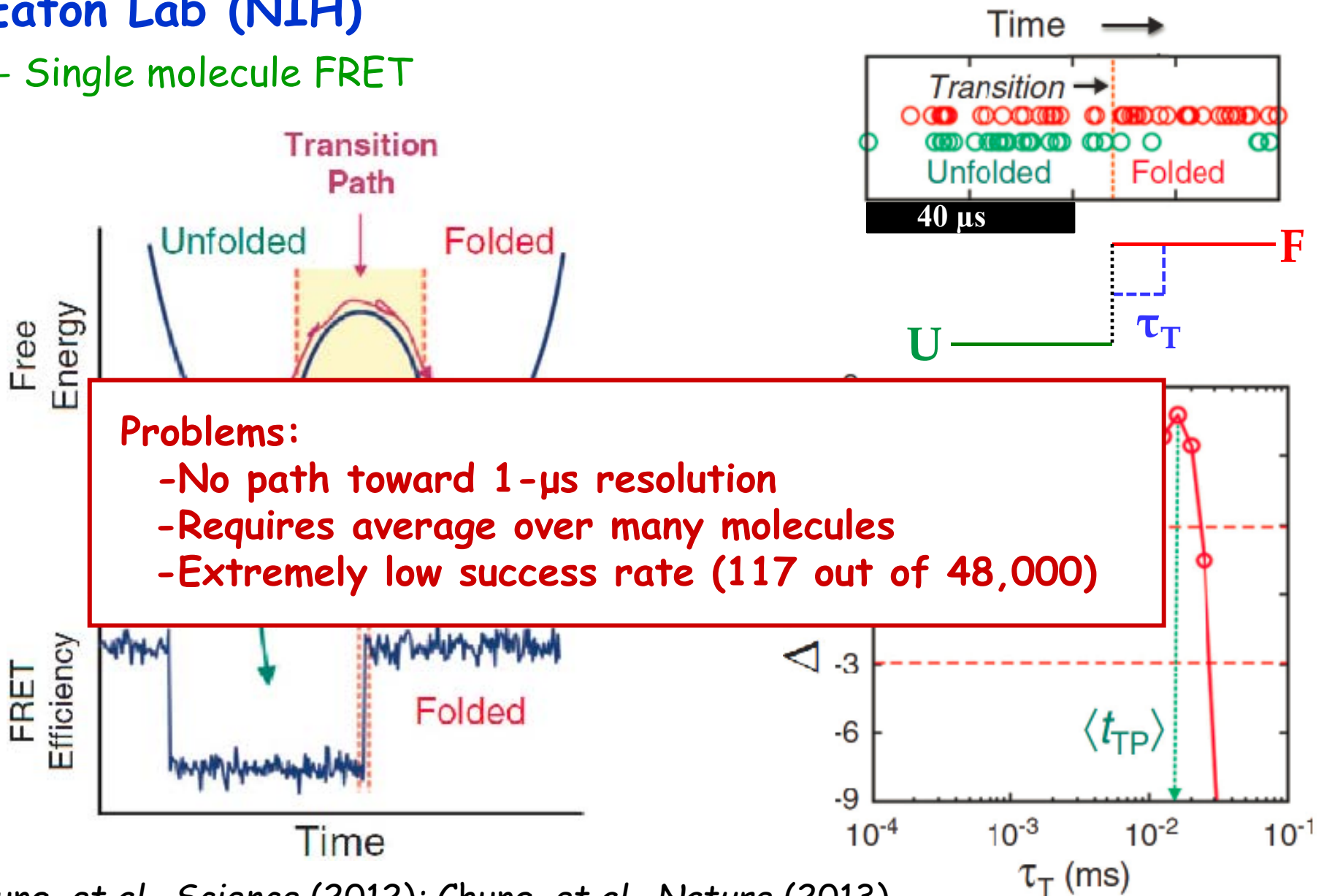
- Goal: Identify states, pathways, and dynamics



Current state of the art: 10- μ s temporal resolution

• Eaton Lab (NIH)

- Single molecule FRET

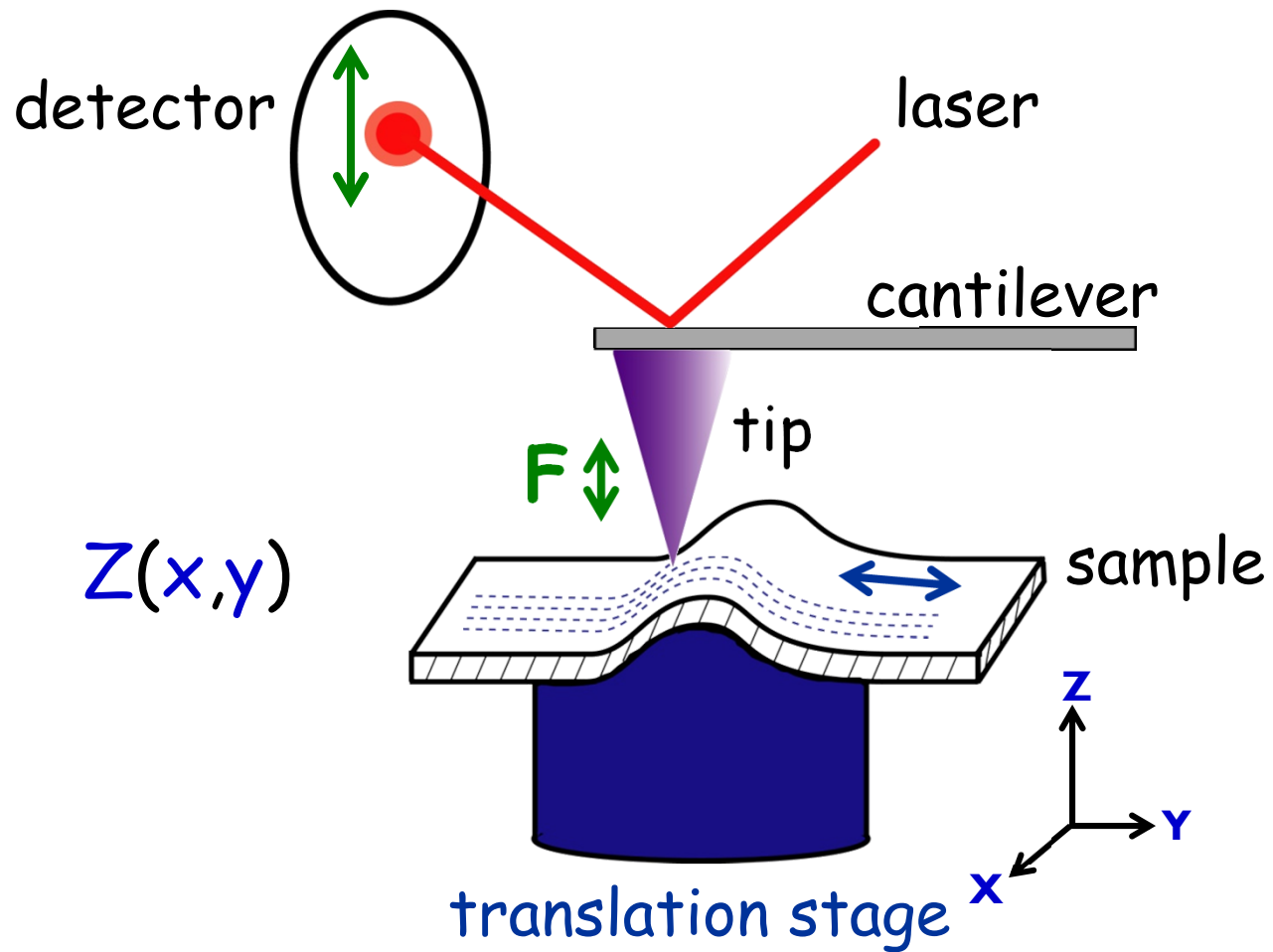


Problems:

- No path toward 1- μ s resolution
- Requires average over many molecules
- Extremely low success rate (117 out of 48,000)

Chung, et al., *Science* (2012); Chung, et al., *Nature* (2013)

Basics of atomic force microscopy



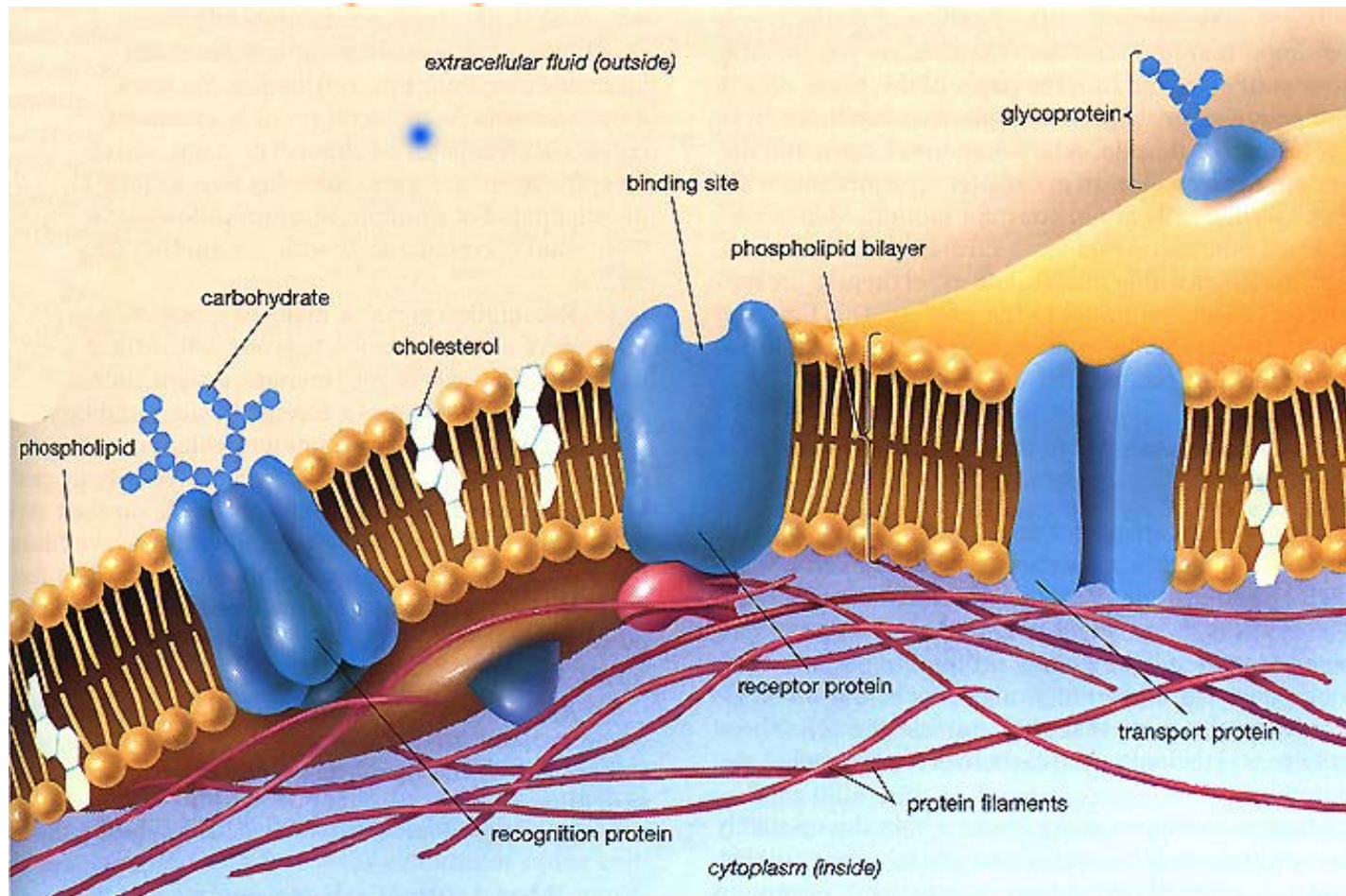
Membrane proteins: a frontier in structural biology

Motivation:

Target for 50% of future drugs
30% of genome
1% of Protein Data Bank

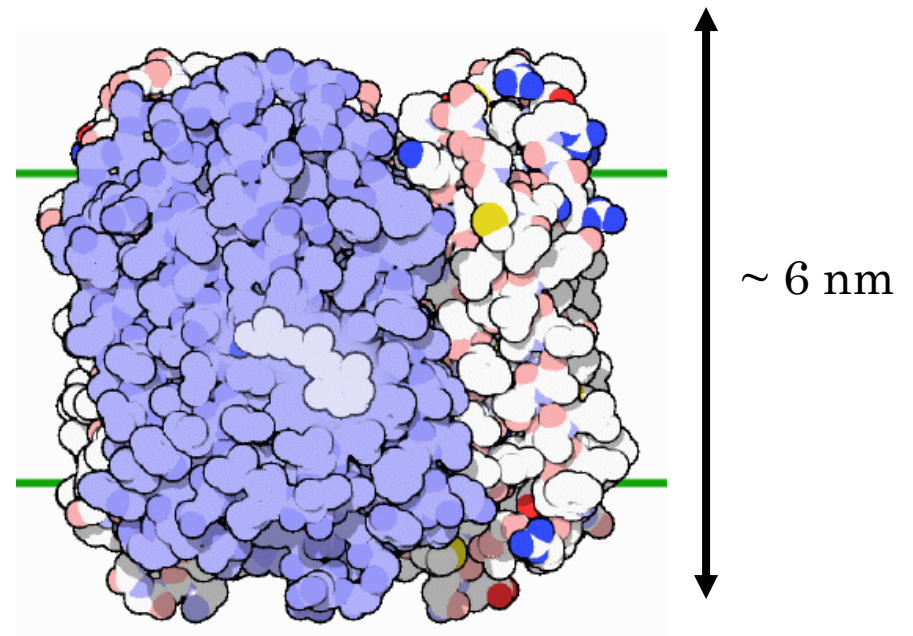
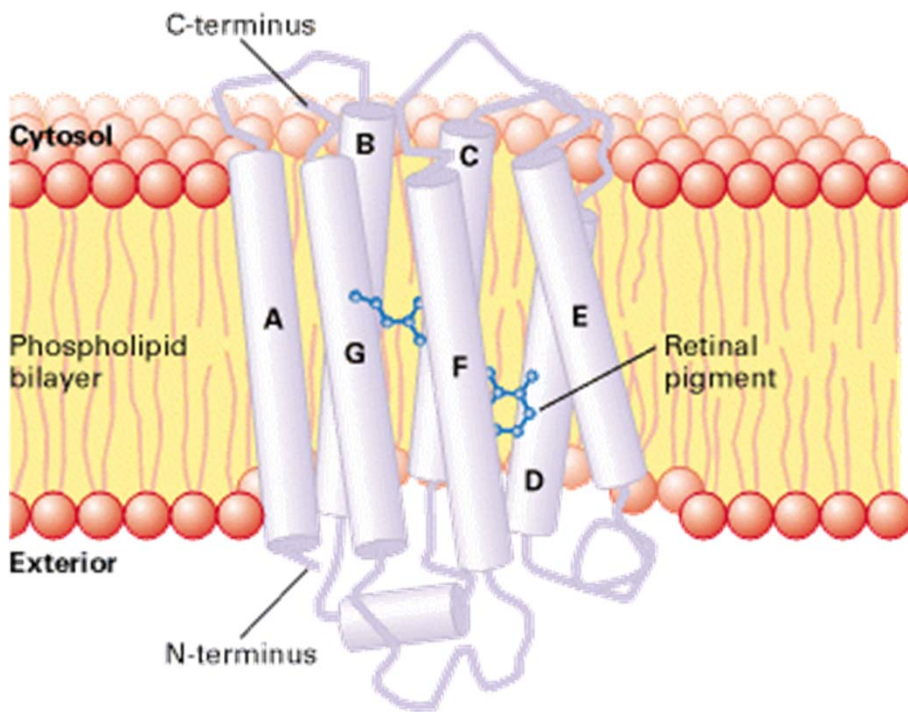
Problem:

Difficult to characterize by:
Crystallography, NMR spectroscopy,
& electron microscopy



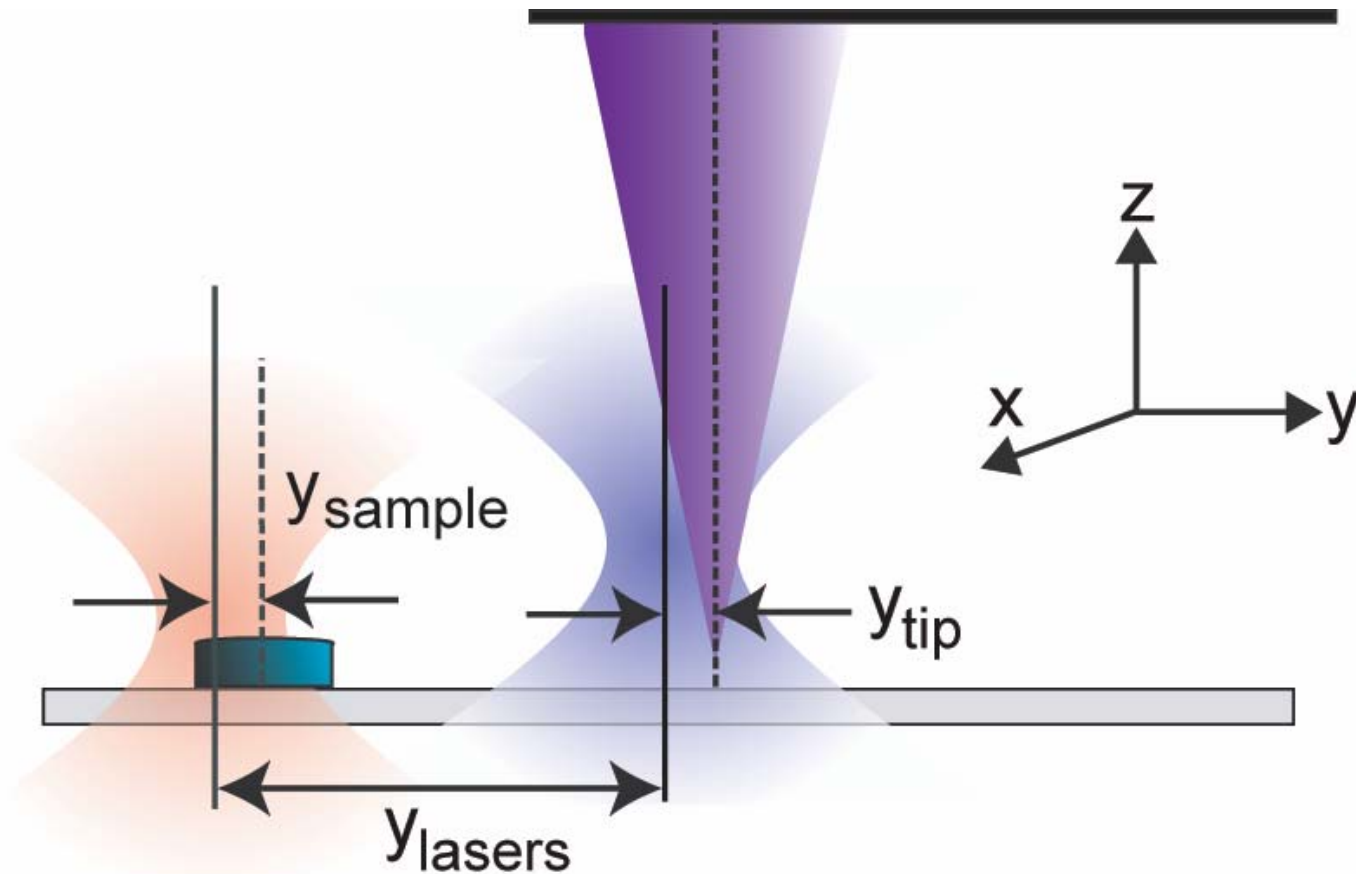
Bacteriorhodopsin is a model membrane protein

- Trimers of BR make “purple membrane”
- Extensively characterized by AFM
(Hansma, Gaub, Engel, Müller, ...)

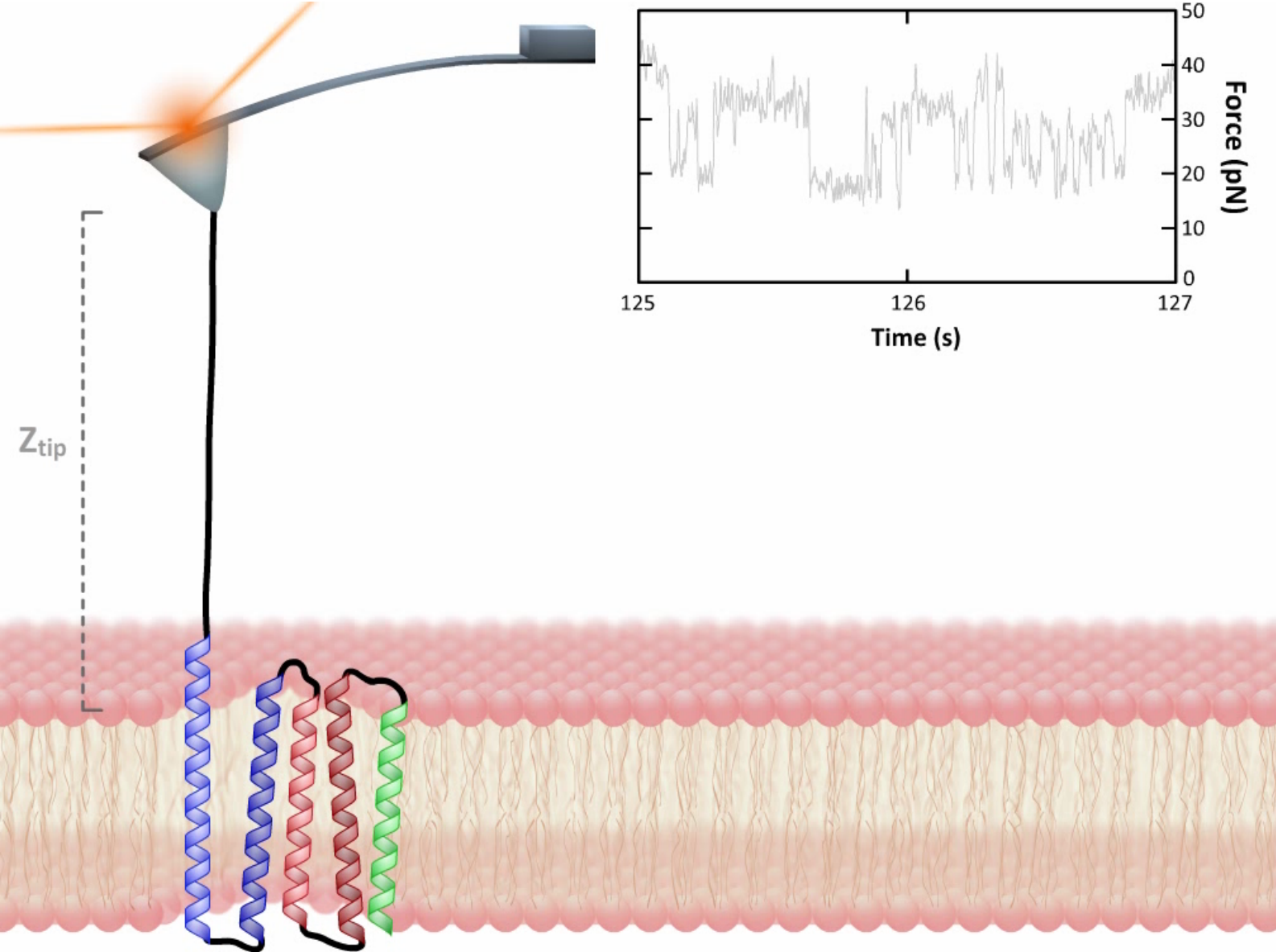


Optically stabilized AFM

- Locally measure
- Actively stabilize



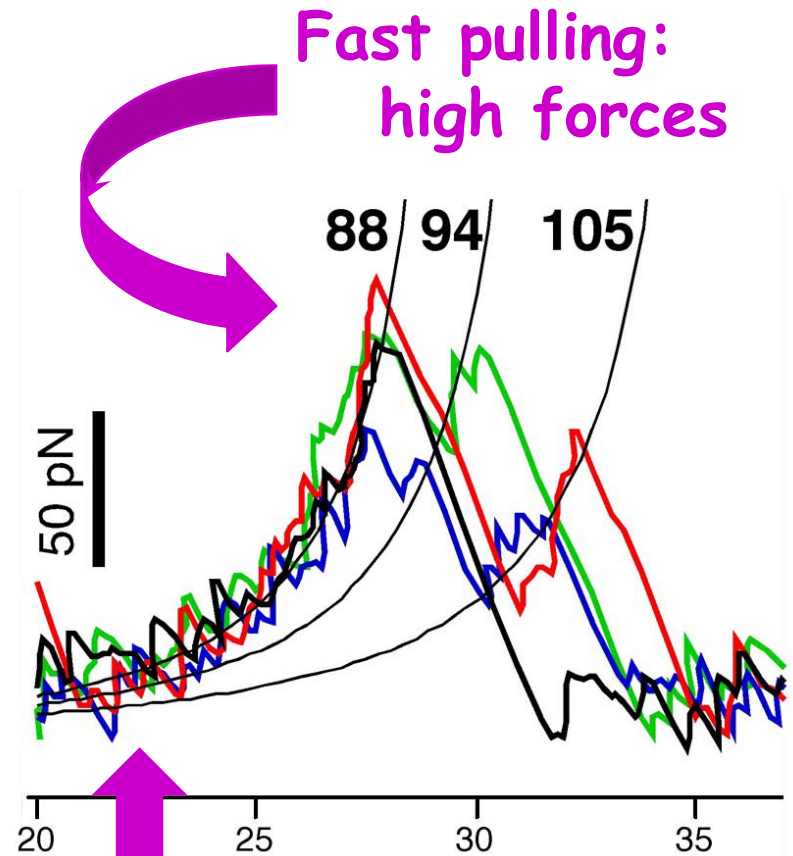
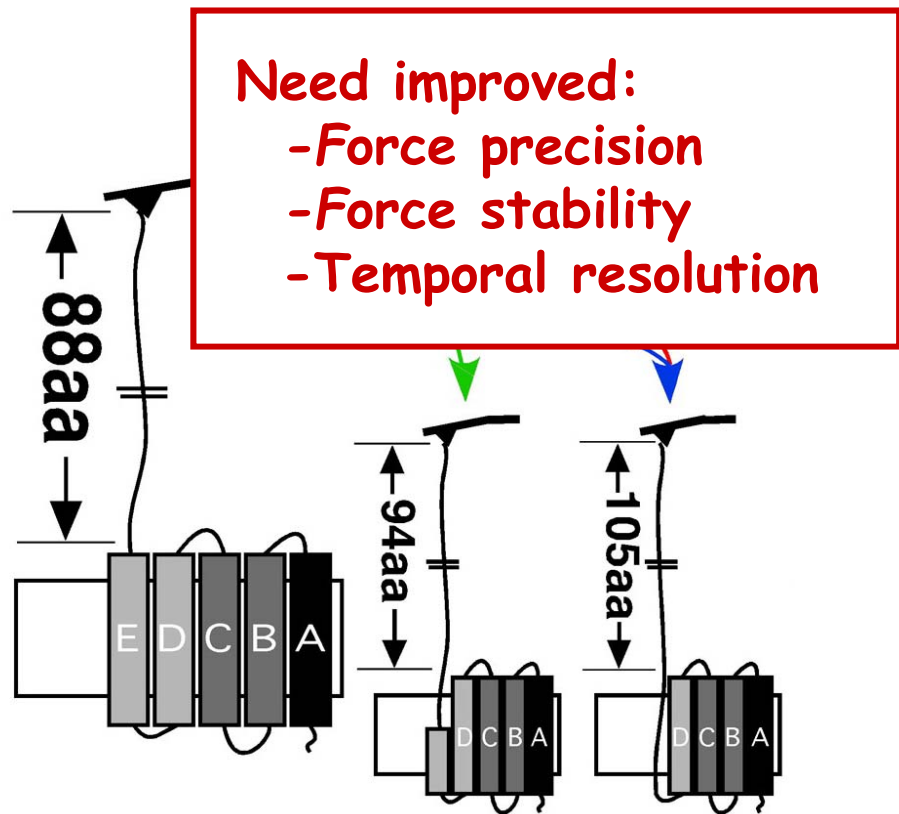
Folding and unfolding of a membrane protein



Challenges to interpretation

- **Unfolding intermediates previous described**

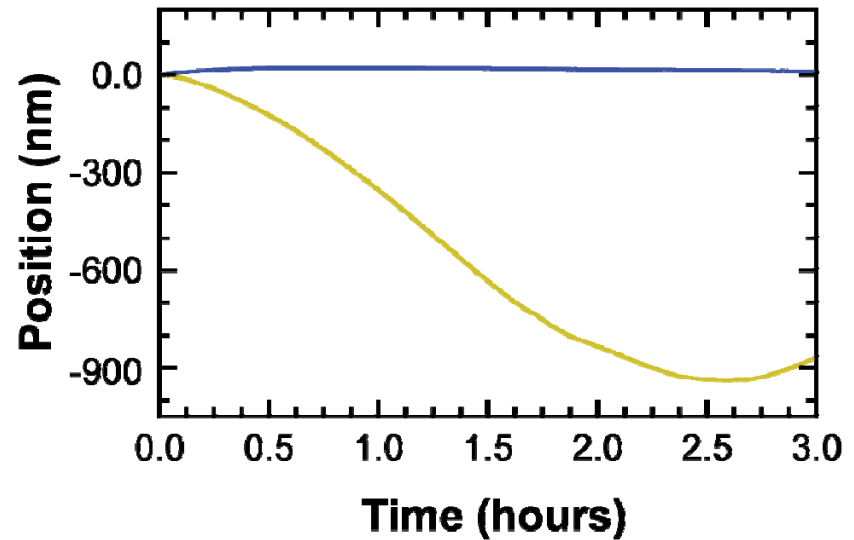
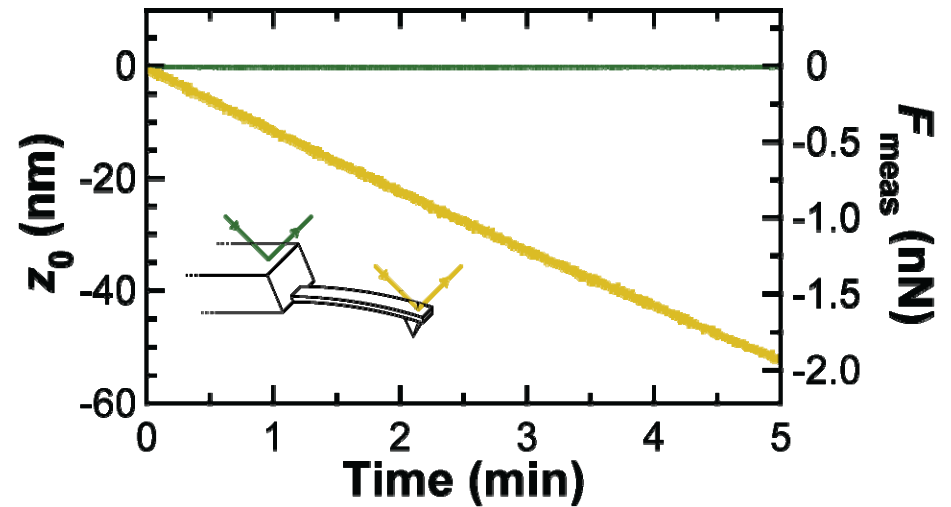
- Gaub and Muller lab



Slow pulling:
low forces

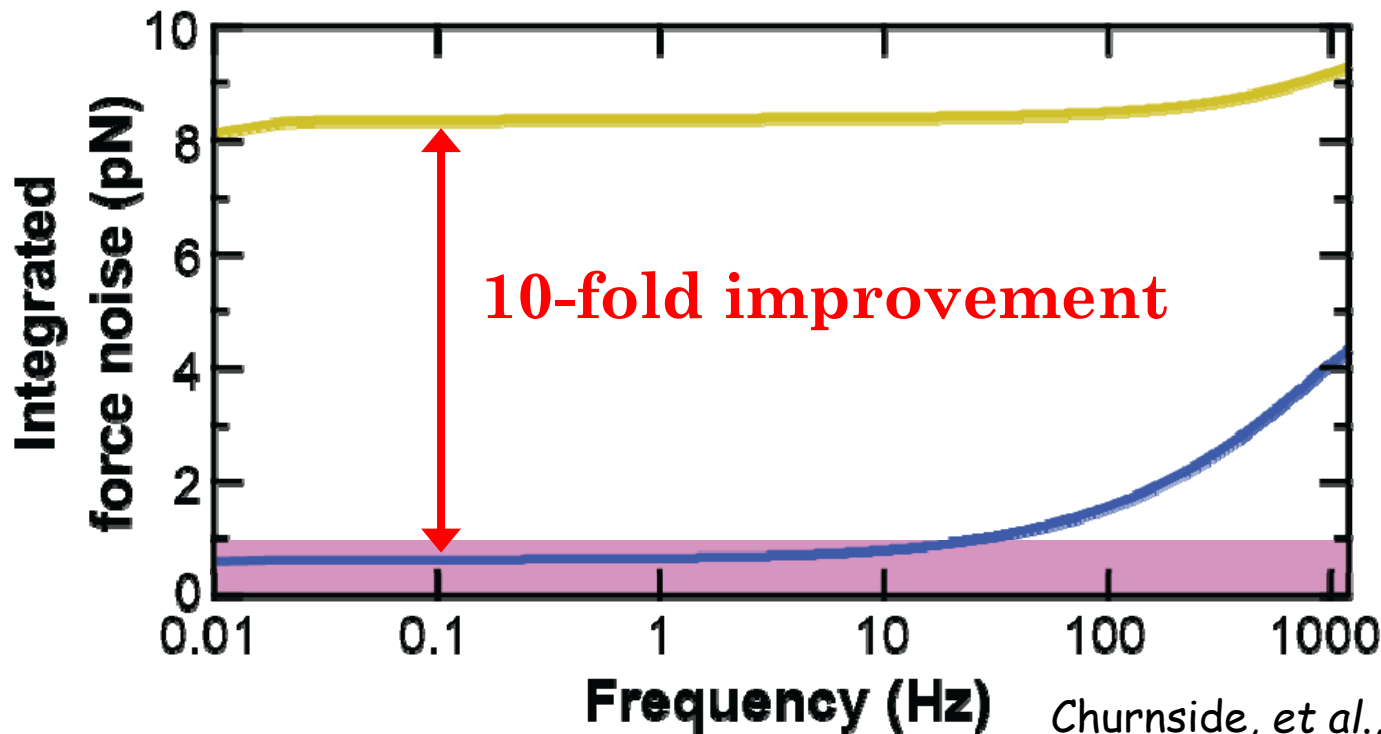
Force drift arises from gold coating

- **Cantilever position drifting**
 - Not external opto-mechanical stability
- **Gold coating causes drift**
 - Removing gold dramatically improves force stability



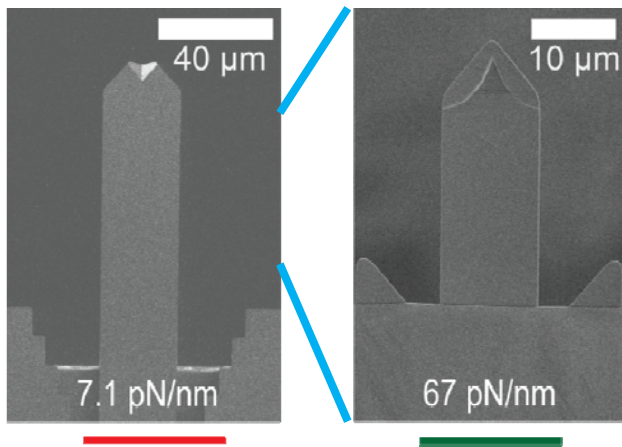
Sub-pN force precision and stability for bioAFM

- **Reflectivity not essential**
 - Commercial and US-AFM
- **Routine**
 - Achieved for 60% of cantilevers tested ($N = 14$)
- **Timely**
 - Achieved 30 min after wetting
 - No long "settling" required



Churnside, et al., *Nano Letters* (2012)

Distinct temporal regimes in AFM force spectroscopy

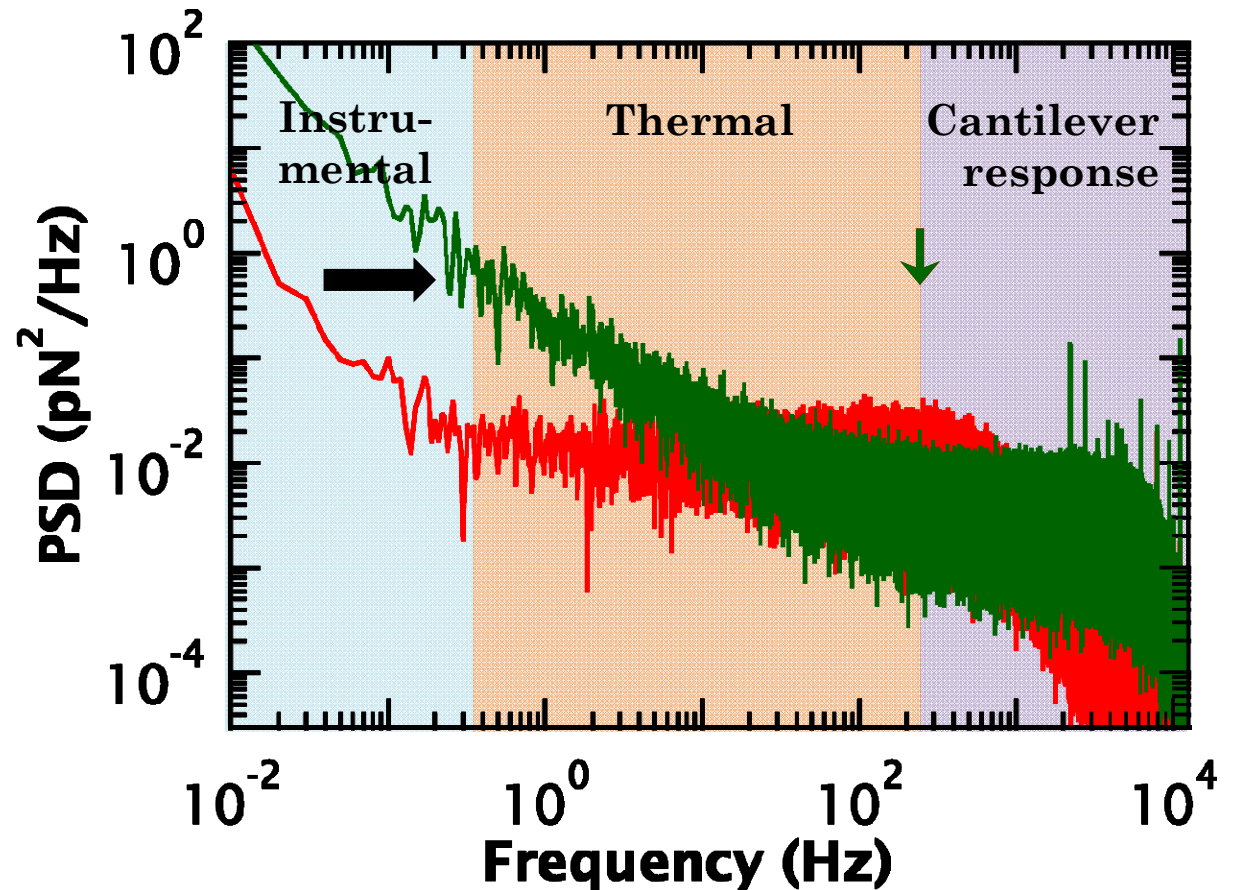


Design goals:

- Soft and short

Challenge:

$$k \propto \frac{wt^3}{l^3}$$

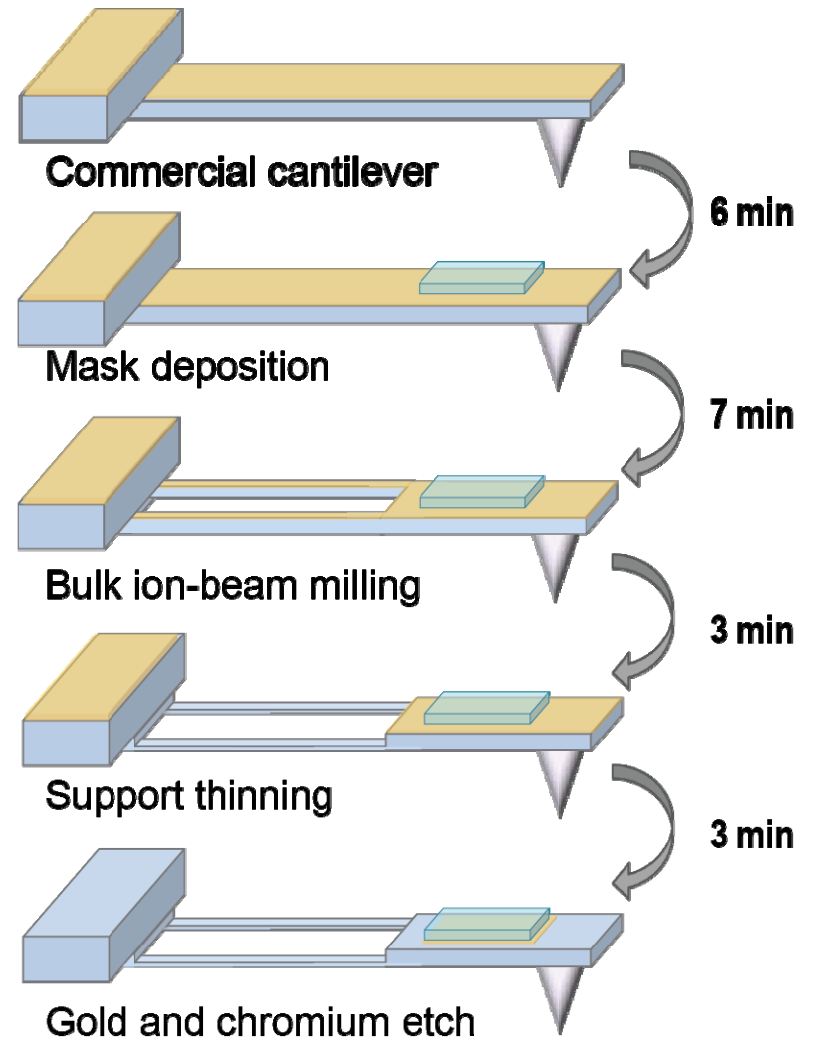
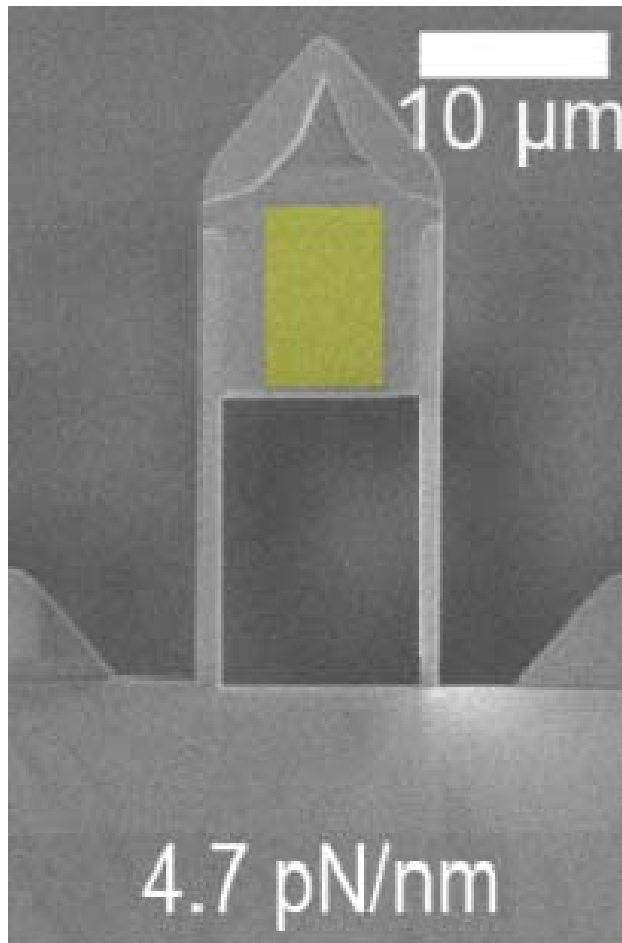


Efficient production of FIB-modified cantilevers

- Ten-fold decrease in hydrodynamic drag and stiffness

- FIB-modified

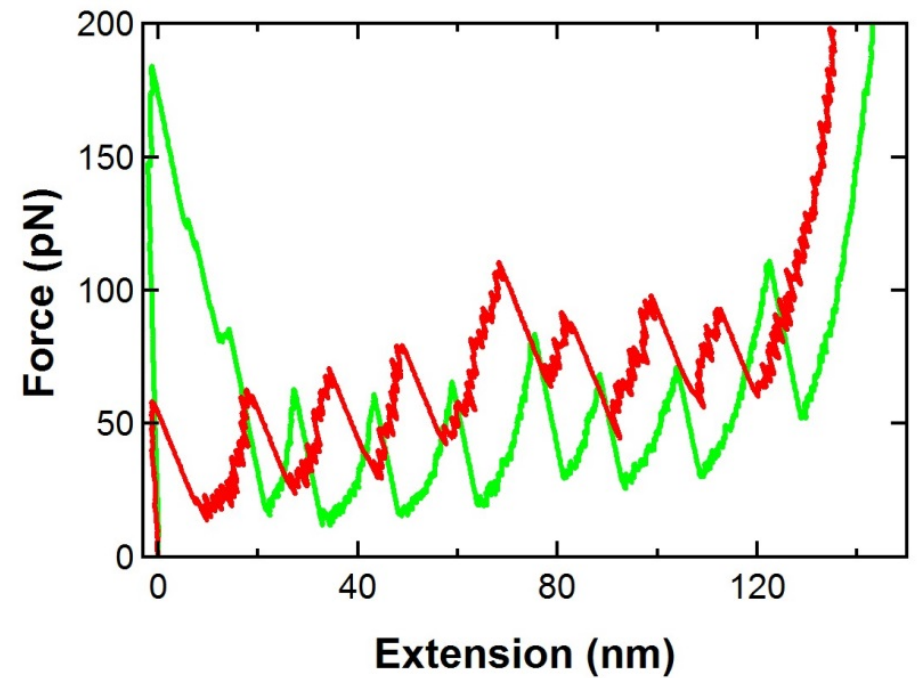
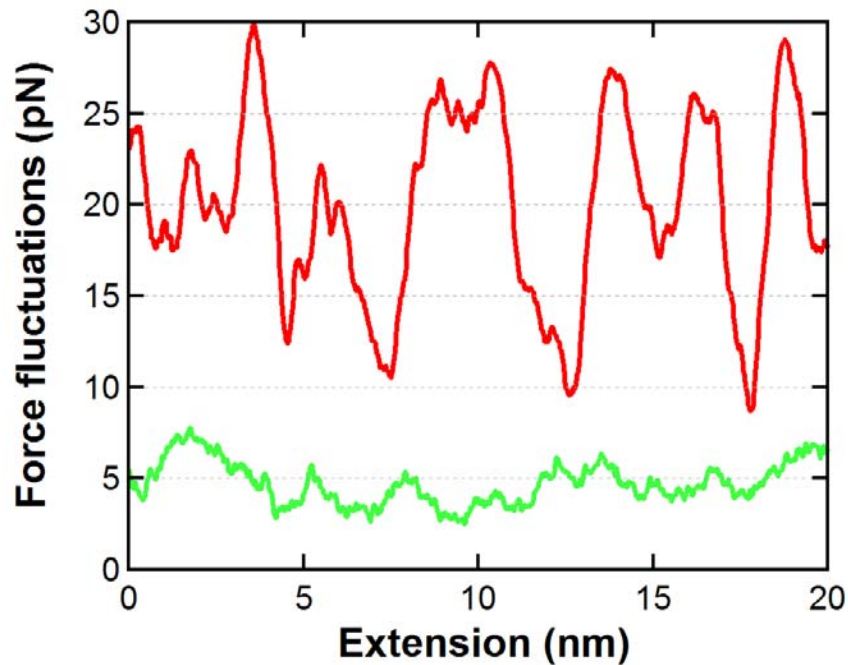
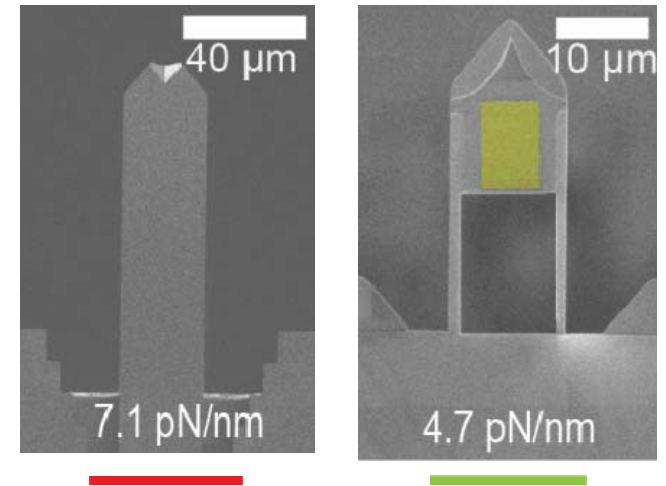
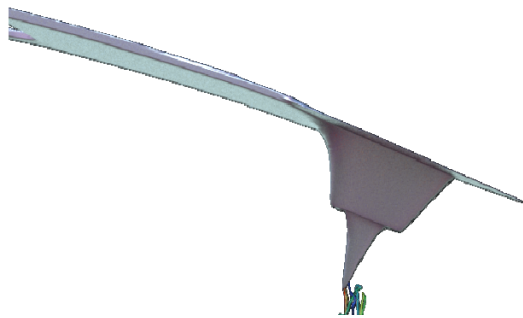
see: Hodges, *Rev. Sci Instrum.* 2001



Bull, *et al.*, *ACS Nano* (2014)

Modified cantilevers improve biophysical data

- **Unfolding a polyprotein**
 - Widely used single-molecule assay

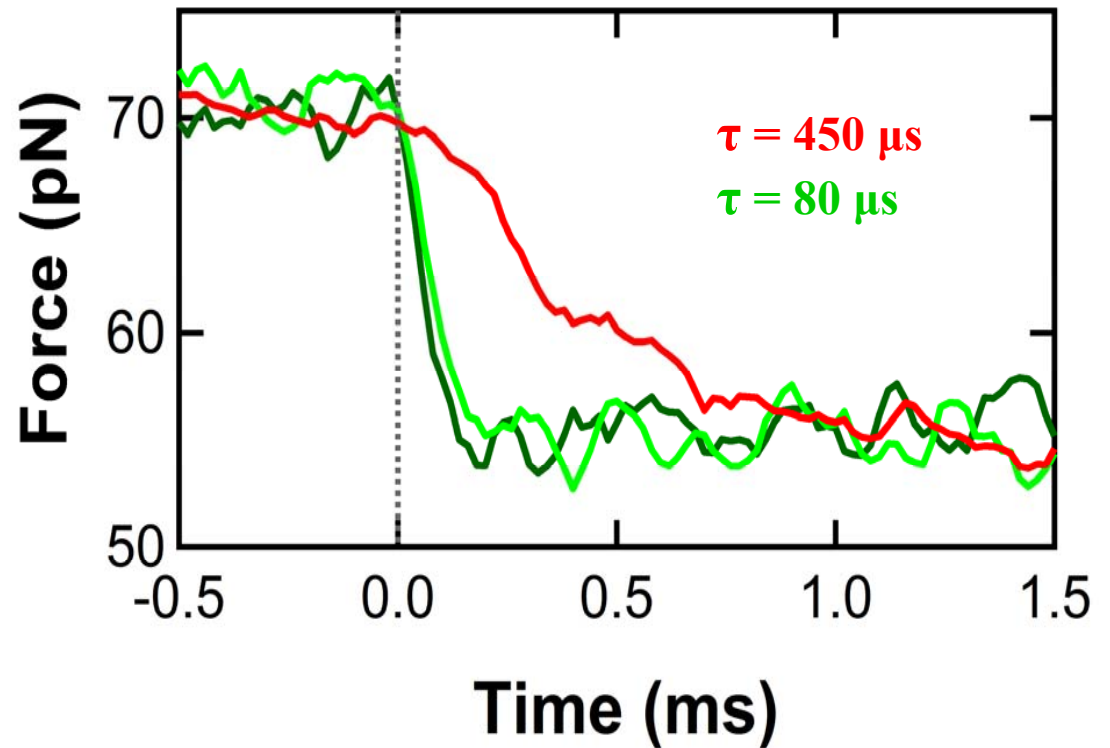
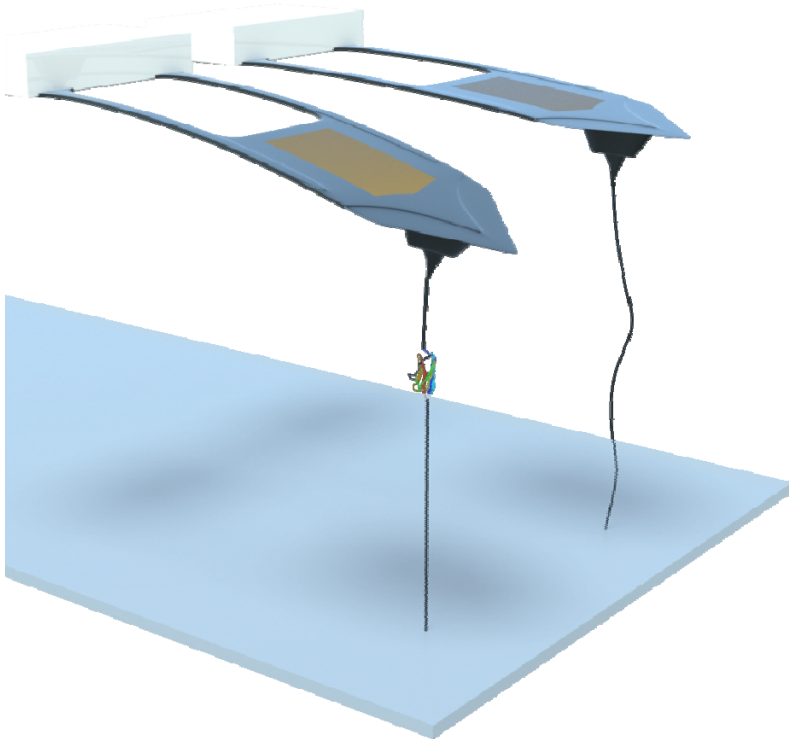


Sensitive but responsive cantilevers

- Measuring the response function

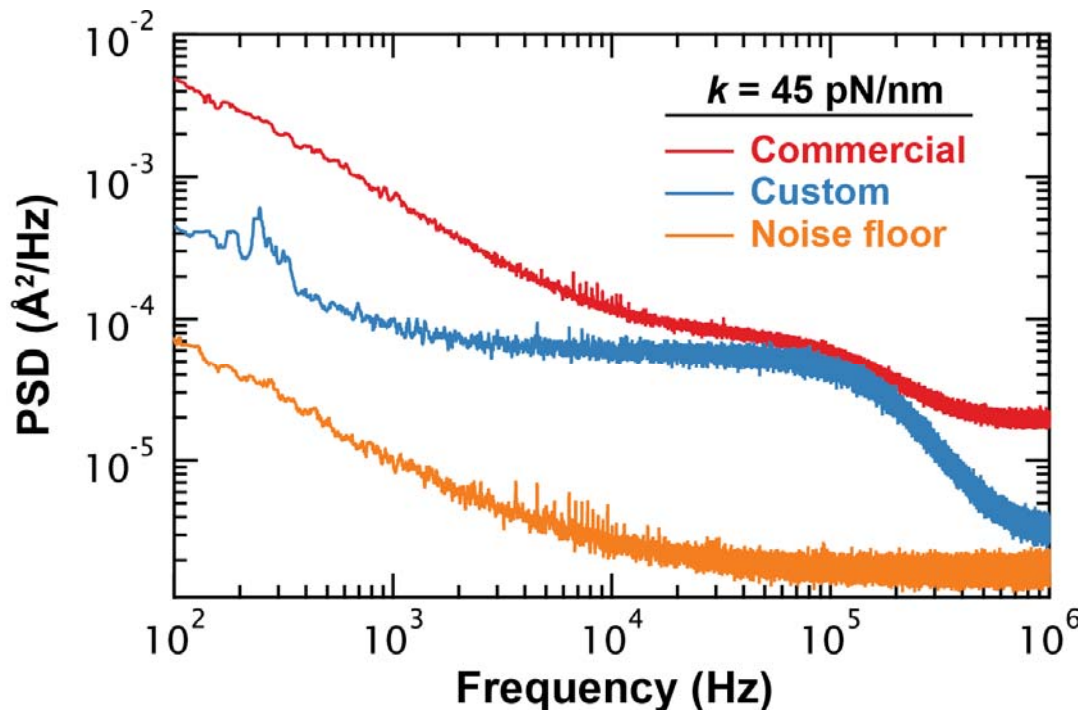
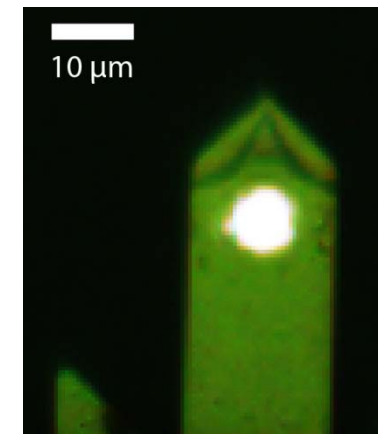
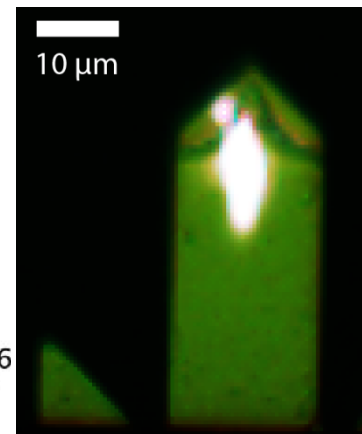
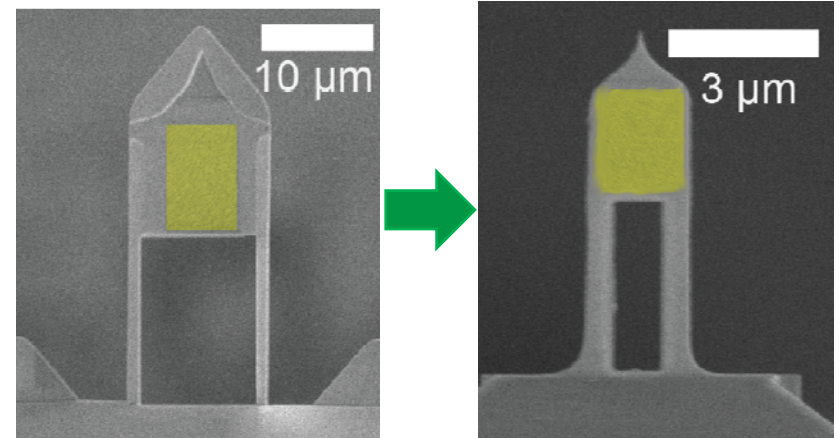
- Fast protein dynamics masked by cantilever response

$$\tau_{\text{lever}} = \frac{\beta_{\text{lever}}}{k_{\text{lever}}}$$



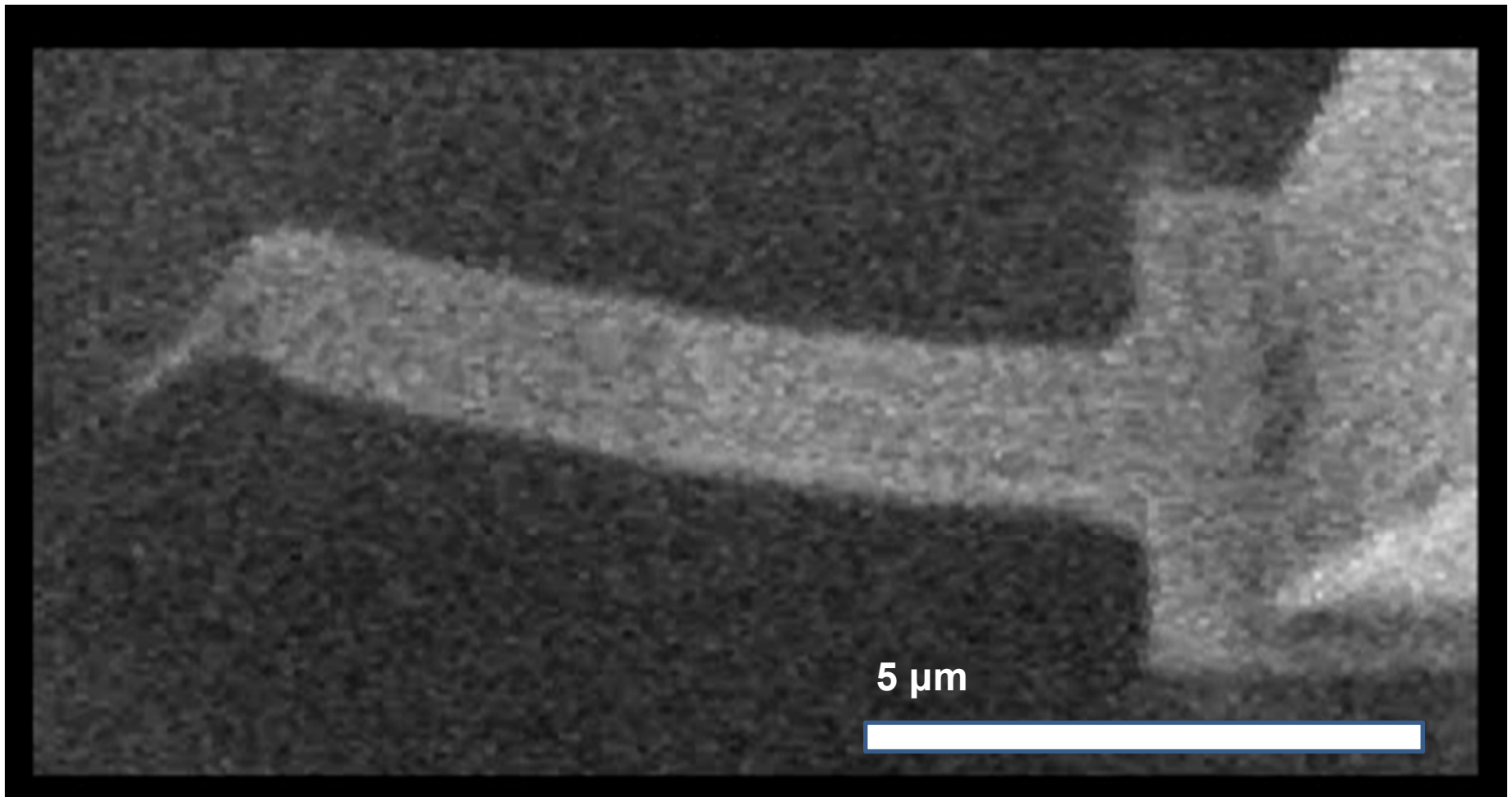
Next step: modifying and detecting ultrashort cantilevers

- Established an efficient fabrication process
 - Compensated for significant bending
- Develop new detection laser
 - Very small spot size ($3\ \mu\text{m}$)
 - Retrofitted into commercial AFM

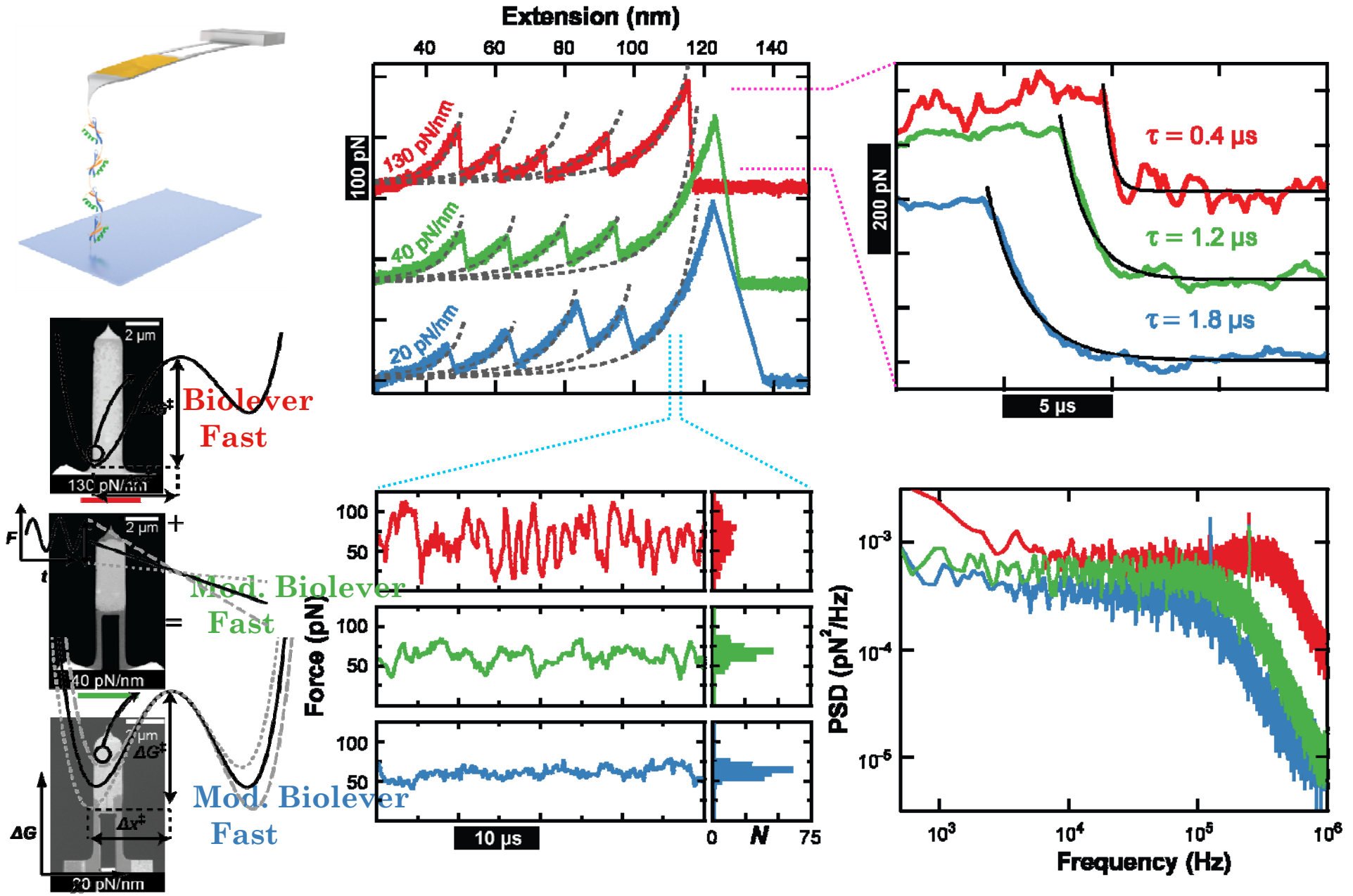


Modification routinely done by skilled undergrads

- Real-time imaging improves yield
 - Thinning bends cantilevers in opposite direction
- Rate: 2-4/hr
 - Limit: handling

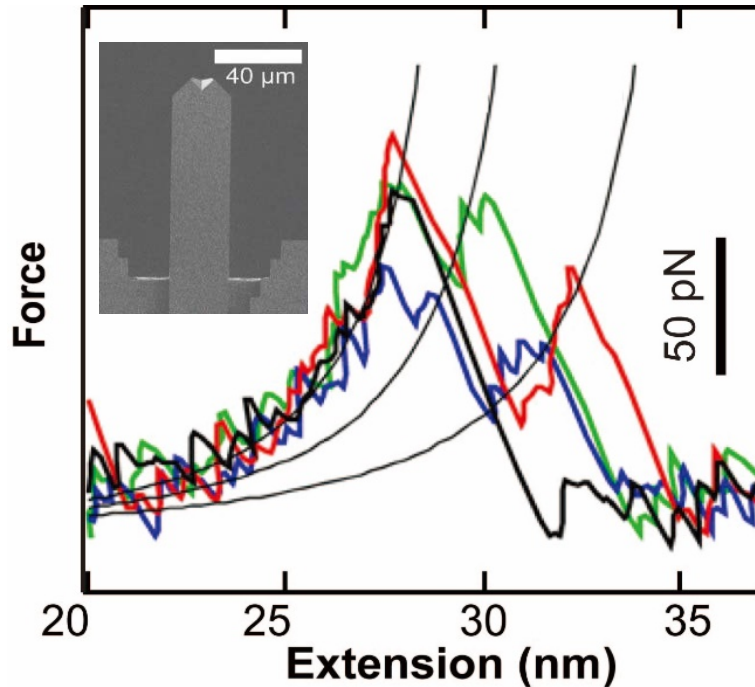


Probing protein unfolding with 1- μ s resolution

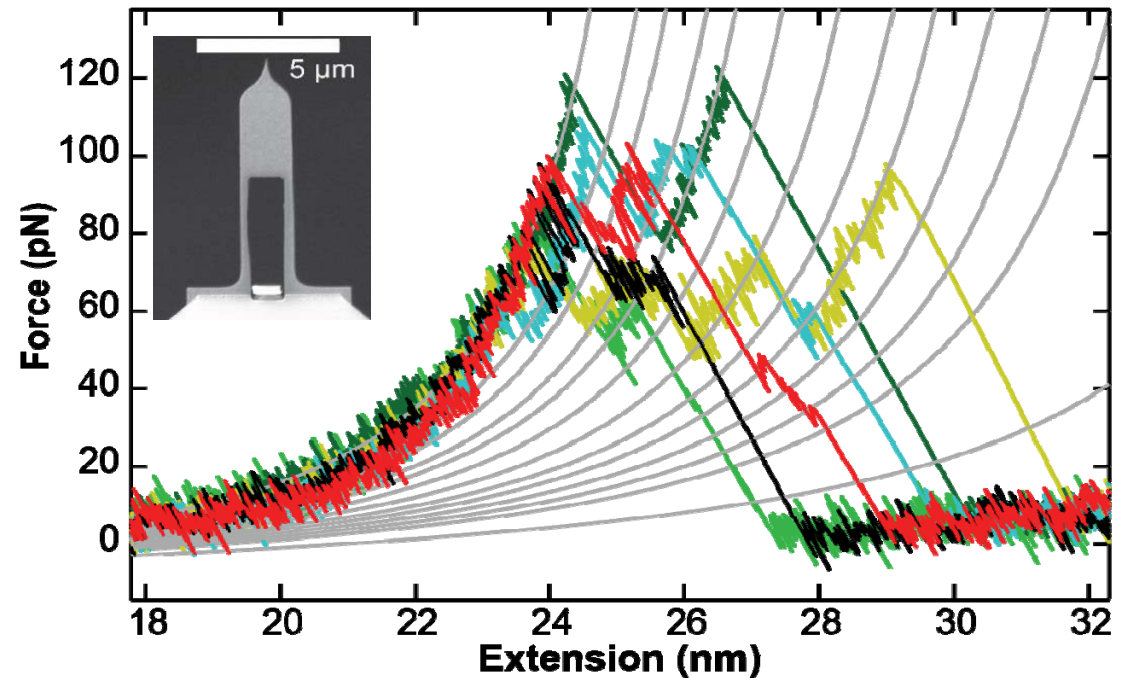


Putting it all together:

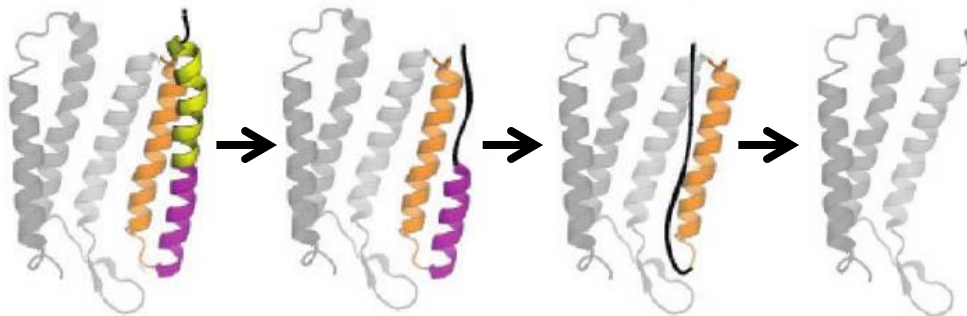
Plethora of new folding intermediates revealed in bR



(Zocher, *ACS Nano*, 2012)



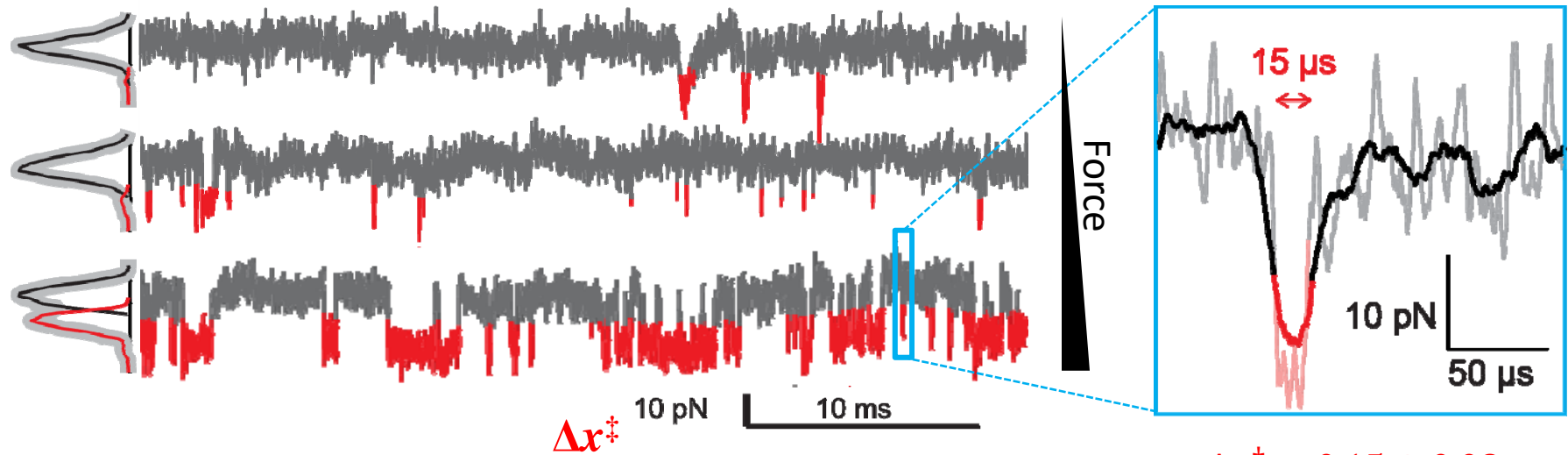
(Perkins lab, unpublished)



• Underlying instrumental advances:

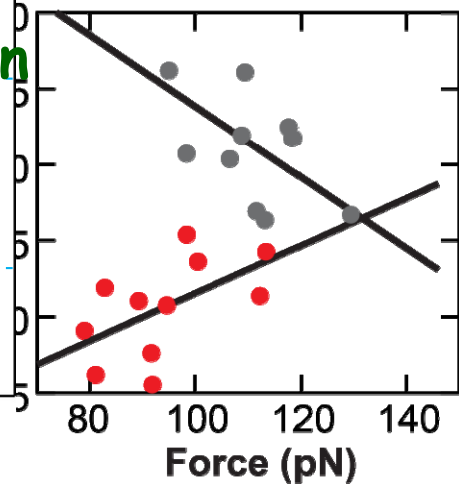
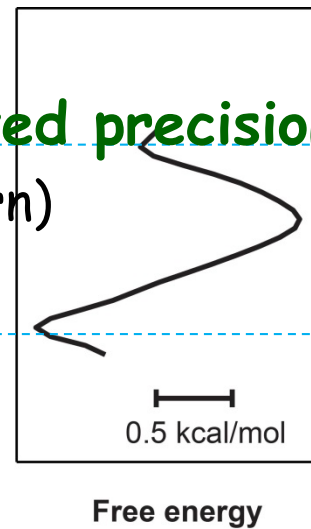
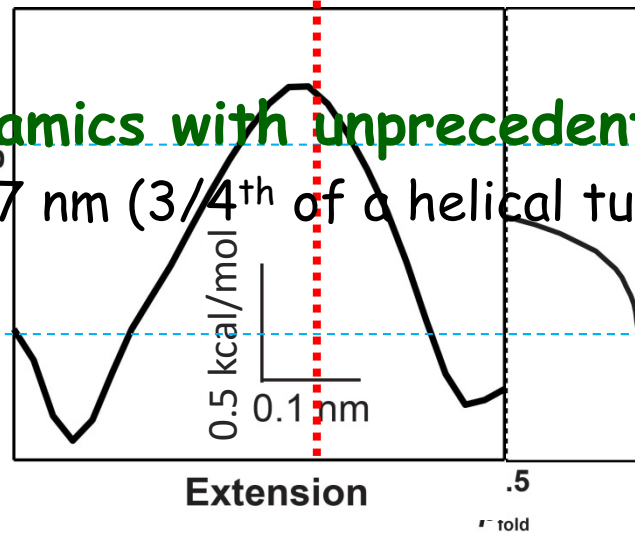
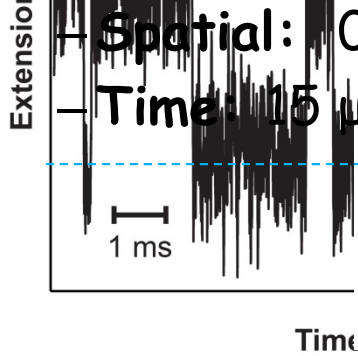
- Time resolution improved: 100-fold
- Force precision improved: 20-fold
- Force stability improved: 10-fold

A new regime for AFM: *equilibrium folding and unfolding*



$$\Delta x^\ddagger = 0.15 \pm 0.08 \text{ nm}$$

• Probing dynamics with unprecedented precision



$$G(x) = \beta^{-1} \ln \left(-D(x) \frac{dp_{fold}}{dx} \right) \quad \text{Chodera \& Pande, PRL, 2011}$$

$$k \propto e^{-F\Delta x^\ddagger/k_B T}$$

High-quality, high-throughput force spectroscopy

- **Current limit: nonspecific attachment**
 - Adsorb biomolecules randomly onto surface
- **Solution: site-specific anchoring**
 - Pursued by many groups

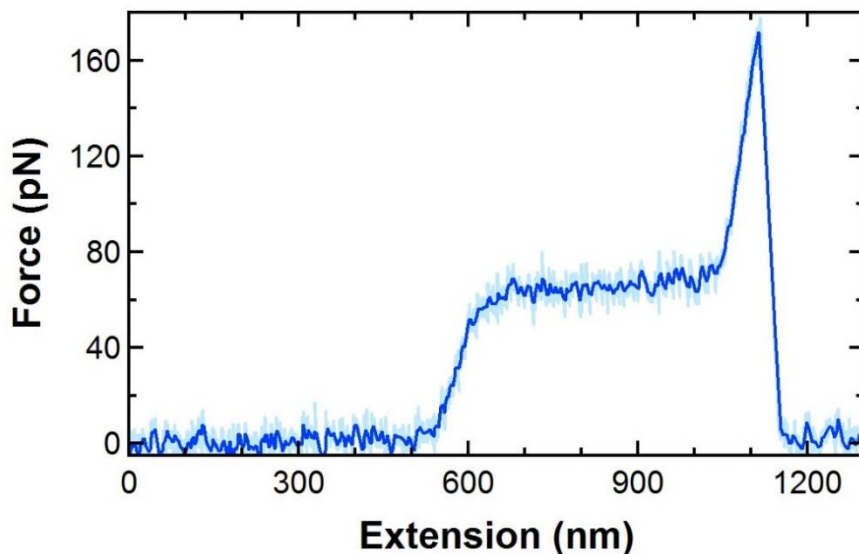
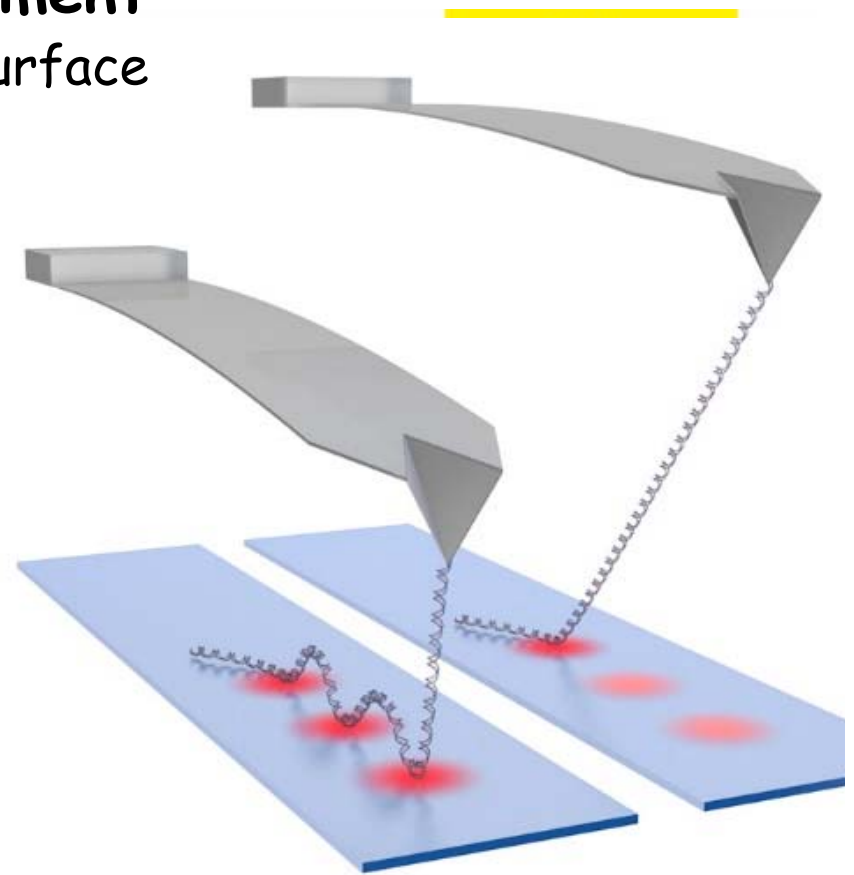
Zimmermann, *et al.*, *Nature Protocols* (2010)

Taniguchi, *et al.*, *Langmuir* (2010)

Stahl, *et al.*, *PNAS* (2012)

Popa, *et al.*, *JACS* (2013)

...



- **Dramatically increased throughput**
 - **Non-specific: 5 HQ traces/month**
 - **Specific: 5 HQ traces/min**

Walder, *et al.*, in preparation

Accelerating studies of diverse proteins

- **Modular construct**

- Covalent surface anchoring
- Reversible tip attachment

■ = NuG2 ● = Protein of Interest

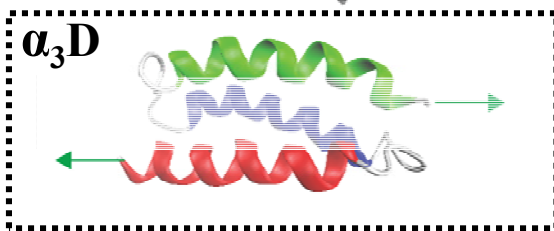


LeBlanc, et al., in preparation

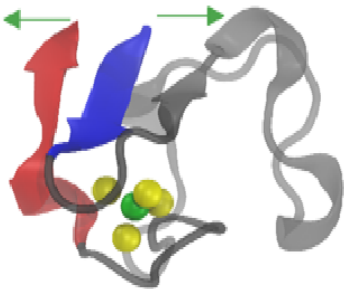
NuG2



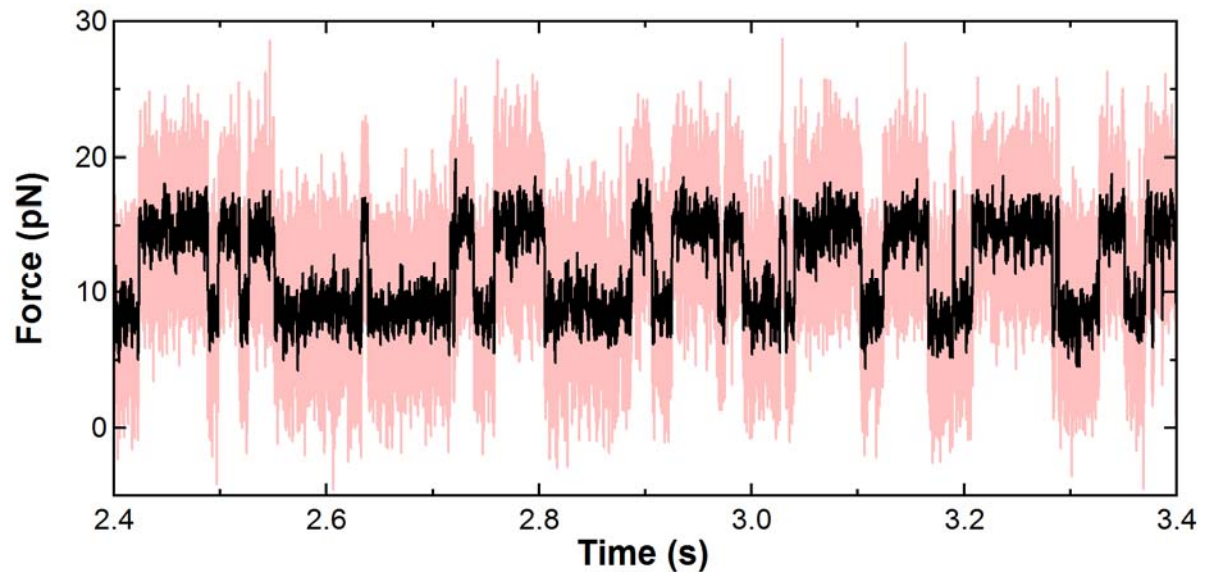
α_3D



Rubredoxin



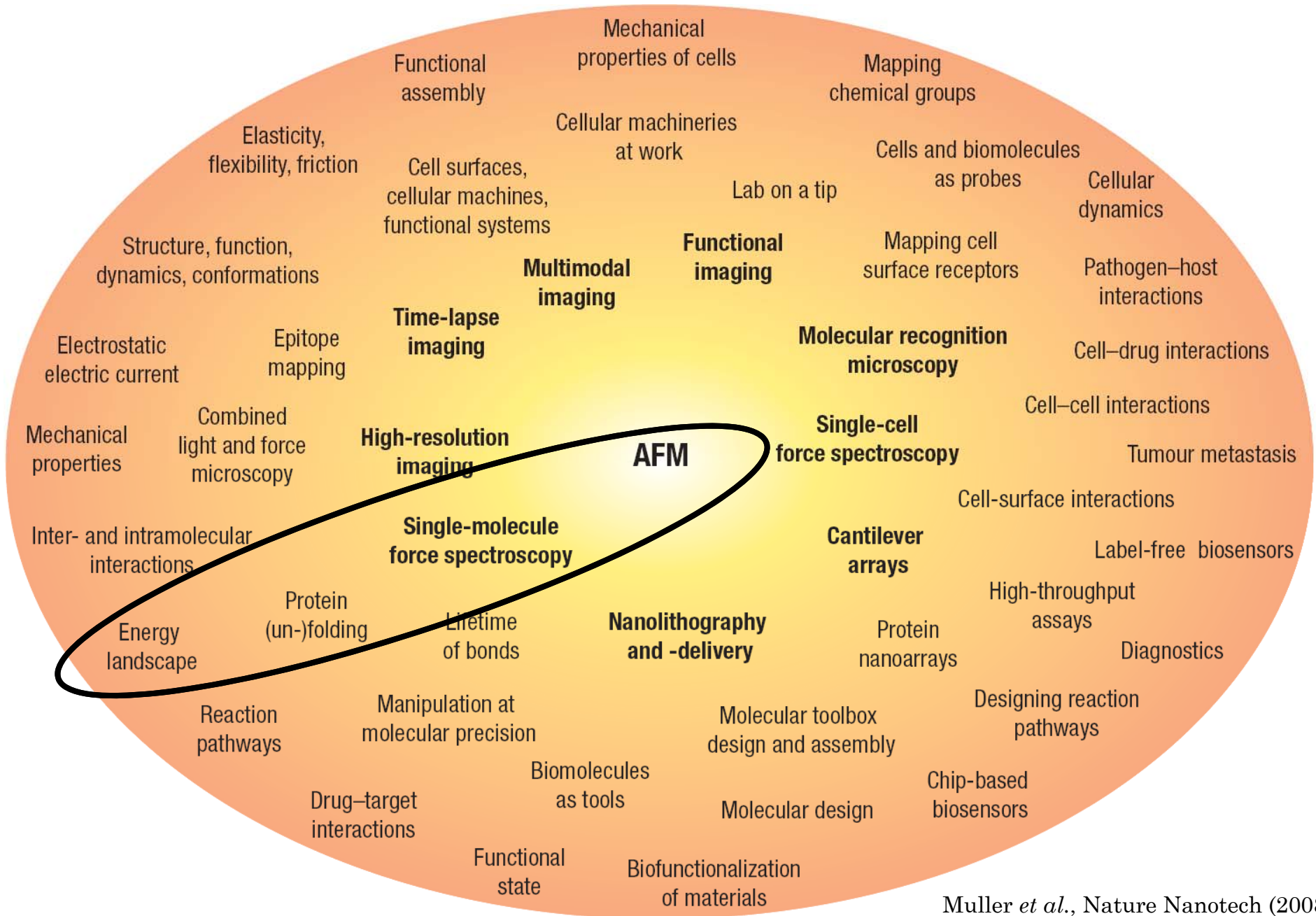
Calmodulin



- Unprecedented precision for AFM

- Metrological advances still needed!

Advances in metrology broadly enable bio-AFM



Acknowledgements

Current Perkins Group:

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